

PROSTAGLANDINS E₁, E₂ AND I₂: EVIDENCE FOR THREE
DISTINCT ACTIONS IN VASCULAR SMOOTH MUSCLEM.S. Manku, D.F. Horrobin, S.C. Cunnane, A.I. Ally, M. Karmazyn,
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Summary: In the perfused mesenteric artery of the rat prostaglandins (PGs) E₁, E₂ and I₂ had distinct actions. PGE₂ potentiated pressor responses to noradrenaline, angiotensin II and potassium ions. PGE₁ potentiated responses to noradrenaline and angiotensin at low concentrations and inhibited them at high concentrations: no concentrations had any effect on potassium responses. PGI₂ inhibited responses to noradrenaline and angiotensin but had no effect on potassium responses. These three distinct actions suggest that the binding sites for the three PGs in this vascular muscle must be distinct.

Introduction

PGE₁ and PGE₂ have many similar actions. Also the recently discovered PGI₂ (prostacyclin) and PGE₁ have similar actions, particularly on adenylate cyclase (1). It is therefore often assumed that there must be considerable similarity between the receptors for the three PGs. We here present evidence that the three PGs have unequivocally distinct effects in rat mesenteric smooth muscle indicating three different receptors.

Methods and Materials

We used the perfused superior mesenteric vascular bed of the male rat as previously described in detail (2,3,4,5). Under ether anesthesia the artery was cannulated and its vascular bed dissected out and mounted in an organ bath. Using a peristaltic pump the preparation was perfused with Krebs-bicarbonate buffer at a flow rate of 3 ml/minute. Perfusion pressure was recorded via a side arm off the arterial cannula. Test injections into another side arm of pressor agents dissolved in 0.1 ml buffer caused brief vasoconstriction and hence elevation of pressure. We have previously shown that both the baseline pressure and the responses to a fixed dose of pressor agent remain stable for many hours (3,4). Actual doses of pressor agents used, each sufficient to give a 40-60% maximal pressor response, were 10 ng noradrenaline as the bitartrate (Sigma), 7 micromol of potassium chloride and 1 ng of angiotensin II (Sigma). The pressor effect of potassium injections is not dependent on release of noradrenaline from nerve endings since it persists even after complete abolition of adrenergic effects by phenoxybenzamine. Responses to potassium and vasopressin are rapidly abolished by use of a calcium-free buffer and presumably depend on stimulation of

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calcium entry from outside. Responses to noradrenaline and angiotensin remain at 60-80% of their original level on removal of extracellular calcium and presumably depend largely on calcium release from intracellular stores (3,4).

We tested the actions of PGE₂, PGE₁, and PGI₂ on pressor responses to noradrenaline (6 experiments each), angiotensin (4 experiments each) and potassium (6 experiments each). Once the preparation had stabilized, three test injections of the pressor agent to be used were given: the mean amplitude of these responses was taken as 100% and subsequent results were expressed as percentages of this. Progressively increasing concentrations of each PG were then added to the buffer, each being present for 10 minutes: at the end of that time the response to the same fixed dose of pressor agent being used was tested. PGE₁ and PGE₂ are stable and were supplied by Upjohn. PGI₂ as its sodium salt was synthesized as previously described (6) from PGF_{2α} methyl ester. This procedure provided a basic ethanolic solution of PGI₂ in a stable form (10⁻³M PGI₂, 8 x 10⁻³M sodium base, 10⁻³M sodium iodide). This solution was directly diluted with the buffer being used as perfusate. The solvent without PGI₂ was similarly diluted and found to have no effects on baseline pressure or on responses to any of the pressor agents. Preliminary experiments showed that the PGI₂ effects were rapid, reached a plateau within five minutes and began to fade at about 15 minutes, probably indicating degradation of the PGI₂. PGE₁ and PGE₂ effects also reached a plateau within 5 minutes and then the effect of each concentration remained unchanged for at least 30 minutes.

Results

None of the PGs had any effect on baseline pressure indicating an absence of direct vasodilating or constricting effects.

PGE₂ caused a progressive increase in response amplitude to injections of potassium and noradrenaline (fig. 1) and angiotensin (not shown). PGE₁ potentiated responses to noradrenaline at low concentrations but above 10⁻¹¹M this potentiation became progressively less and was converted to an inhibition above 10⁻⁷M (fig. 2). PGE₁ effects on angiotensin responses were indistinguishable from those on noradrenaline, but PGE₁ failed to alter potassium responses.

PGI₂ failed to change responses to potassium but caused a progressive inhibition of responses to noradrenaline (fig. 3). In six preparations each, before adding the increasing concentrations of PGI₂, either 10 ng/ml PGE₂ or 10 μg/ml indomethacin was added to the buffer and left there while the PGI₂ dose response curve was carried out. The PGE₂ increased responses to about 150% of those obtained with buffer alone. These increased responses were taken as the 100% starting values for the PGI₂ curve. The presence of

Table I. Inhibitory potencies of Leu-enkephalin and fluorescent peptides on the contractions of guinea-pig ileum and on the binding of ^3H -Leu-enkephalin by membranes from mouse striatum.

Compound	Guinea-pig ileum		^3H -Leu-enkephalin
	IC_{50} (nM)	Relative Potency	binding Ki (nM)
Leu-E	67 ± 12	100	5 ± 1
Met-E-(CH_2) ₂ dansyl	50 ± 11	86	10 ± 3
D-Ala ₂ -Met-E-(CH_2) ₂ dansyl	27 ± 4	147	5 ± 2
Met-E-(CH_2) ₅ dansyl	288 ± 15	13	21 ± 5
N-dansyl-Met-E	$1,306 \pm 328$	4	50 ± 25

for β -adrenergic receptors (17). The fine analysis of energy transfer within such molecules in solution can provide interesting data regarding their preferred conformation (18) and could be compared to those provided by other approaches (19,20,21). In addition, the energy transfer between the Tyr and the dansyl groups of the molecule provides an indirect index of the integrity of the molecule : since the biological inactivation consists primarily in the hydrolysis of the Tyr-Gly bond (22), Met-E-(CH_2)₂ dansyl can be used as a substrate for the peptidases involved in this cleavage and, monitoring the *in vitro* changes in fluorescence, provides a convenient mean to evaluate the activity of these enzymes and the actions of inhibitors.

Finally, and perhaps more interestingly, the analysis of energy transfer between tryptophane residues of the receptor molecule and the dansyl group of the probe (8,23) or between Tyr and dansyl chromophores within the latter, could provide interesting data regarding the conformational changes possibly occurring during attachment of the peptide to its recognition sites. Such an approach can be extended to the different endorphins which contain only one Tyr residue at the N-terminal part of the peptide.

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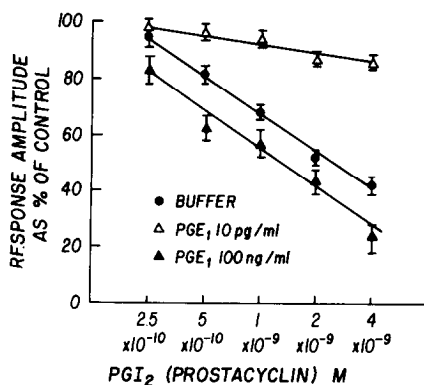


Fig. 4. Changes in response to fixed doses of noradrenaline produced by PGI₂ in plain buffer, and buffer containing 10 pg/ml (2.8×10^{-11} M) PGE₁ or 100 ng/ml (2.8×10^{-7} M) PGE₁. Each point represents the mean \pm SEM for 6 experiments.

PGE₂ failed to alter the PGI₂ effects (fig. 3). Similarly indomethacin which reduced responses to noradrenaline to about 40% of their amplitude in buffer alone (3) failed to modify the PGI₂ curve (fig. 3).

In contrast to these results, PGE₁ did change the PGI₂ effects. 10 pg/ml (2.8×10^{-11} M) PGE₁, the concentration which produces a maximum potentiating effect, markedly attenuated the PGI₂ effect when the experiments were performed as described for the PGE₂/PGI₂ interaction. In contrast, 100 ng/ml (2.8×10^{-7} M) PGE₁ moved the PGI₂ curve slightly to the left (fig. 4).

Discussion

The three PGs have clearly distinct actions on pressor responses in this preparation indicating that three different receptor sites are involved. PGE₂ had equipotent effects on responses dependent on extracellular and intracellular calcium whereas PGE₁ and PGI₂ modified only responses to noradrenaline and angiotensin which seem to be largely dependent on intracellular calcium release. This does not necessarily mean that these PGs act intracellularly because intracellular calcium release could be changed by plasma membrane receptors.

The PGI₂ effects were unaltered by either exogenous PGE₂ or by reduction of endogenous PG synthesis by indomethacin suggesting that they were not

dependent on antagonism of action of an endogenous PG. They were attenuated by a very low concentration of PGE_1 indicating that PGE_1 and PGI_2 may oppose one another's effect on intracellular calcium release. PGE_1 altered the slope of the dose response curve indicating that the interaction between the two PGs was either due to an allosteric mechanism or to a physiological antagonism rather than to competition for the same receptor site. In contrast, the high PGE_1 concentration which itself was weakly inhibitory moved the PGI_2 curve slightly to the left but apparently in parallel. This might possibly be due to activation of PGI_2 receptors by high concentrations of PGE_1 or to stimulation of PGI_2 synthesis or to some other unknown mechanism.

We conclude that the different effects of the three PGs are probably related to their ability to activate specific receptors selectively.

Acknowledgments

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